Transit station or destination? Attendance patterns, movements and abundance estimate of humpback whales off west South Africa from photographic and genotypic matching

J Barendse, PB Best, M Thornton, SH Elwen, HC Rosenbaum, I Carvalho, C Pomilla, TJQ Collins, MA Meeyer & RH Leeney

Mammal Research Institute, University of Pretoria, c/o Iziko South African Museum, PO Box 61, Cape Town, 8000, South Africa
Ocean Giants Program, Wildlife Conservation Society, 2300 Southern Boulevard, Bronx, NY, 10460-1099, USA
Sackler Institute for Comparative Genomics, American Museum of Natural History, Central Park West at 79th Street, New York, NY, 10024, USA
Environment Society of Oman, PO Box 3955, PC 112, Ruwi, Sultanate of Oman
Faculdade de Ciências do Mar e Ambiente - Universidade do Algarve, Campus Gambelas, 8000-139, Faro, Portugal
Oceans and Coasts, Department of Environmental Affairs, Private Bag X2, Rogge Bay, 8012, South Africa
School of Marine Science and Engineering, Portland Square, University of Plymouth, Drake Circus, Plymouth, PL4 8AA, Devon, UK

Available online: 11 Jan 2012

To cite this article: J Barendse, PB Best, M Thornton, SH Elwen, HC Rosenbaum, I Carvalho, C Pomilla, TJQ Collins, MA Meeyer & RH Leeney (2011): Transit station or destination? Attendance patterns, movements and abundance estimate of humpback whales off west South Africa from photographic and genotypic matching, African Journal of Marine Science, 33:3, 353-373

To link to this article: http://dx.doi.org/10.2989/1814232X.2011.637343

PLEASE SCROLL DOWN FOR ARTICLE

Full terms and conditions of use: http://www.tandfonline.com/page/terms-and-conditions

This article may be used for research, teaching, and private study purposes. Any substantial or systematic reproduction, redistribution, reselling, loan, sub-licensing, systematic supply, or distribution in any form to anyone is expressly forbidden.

The publisher does not give any warranty express or implied or make any representation that the contents will be complete or accurate or up to date. The accuracy of any instructions, formulae, and drug doses should be independently verified with primary sources. The publisher shall not be liable for any loss, actions,
claims, proceedings, demand, or costs or damages whatsoever or howsoever caused arising directly or indirectly in connection with or arising out of the use of this material.
Transit station or destination? Attendance patterns, movements and abundance estimate of humpback whales off west South Africa from photographic and genotypic matching

J Barendse1*, PB Best1, M Thornton1, SH Elwen1, HC Rosenbaum2,3, I Carvalho2,3,5, C Pomilla3, TJQ Collins2,4, MA Meyer6 and RH Leeney7

1 Mammal Research Institute, University of Pretoria, c/o Iziko South African Museum, PO Box 61, Cape Town 8000, South Africa
2 Ocean Giants Program, Wildlife Conservation Society, 2300 Southern Boulevard, Bronx, NY 10460-1099, USA
3 Sackler Institute for Comparative Genomics, American Museum of Natural History, Central Park West at 79th Street, New York, NY 10024, USA
4 Environment Society of Oman, PO Box 3955, PC 112, Ruwi, Sultanate of Oman
5 Faculdade de Ciências do Mar e Ambiente – Universidade do Algarve, Campus Gambelas, 8000-139, Faro, Portugal
6 Oceans and Coasts, Department of Environmental Affairs, Private Bag X2, Rogge Bay 8012, South Africa
7 School of Marine Science and Engineering, Portland Square, University of Plymouth, Drake Circus, Plymouth PL4 8AA, Devon, UK

* Corresponding author, e-mail: jaco.barendse@gmail.com

Manuscript received February 2011; accepted August 2011

Humpback whales Megaptera novaeangliae found off west South Africa (WSA) are known to display an atypical migration that may include temporary residency and feeding during spring and summer. At a regional scale there is uncertainty about how these whales relate to the greater West African Breeding Stock B as a whole, with evidence both for and against its division into two substocks. A database containing sighting information of humpback whales intercepted by boat in the WSA region from 1983 to 2008 was compiled. It included a total of 1 820 identification images of ventral tail flukes and lateral views of dorsal fins. After systematic within- and between-year matching of images of usable quality, it yielded 154 different individuals identified by tail flukes (TF), 230 by left dorsal fins (LDF), and 237 by right dorsal fins (RDF). Microsatellite (MS) matching of 216 skin biopsies yielded 156 individuals. By linking all possible sightings of the same individuals using all available identification features, the periodicity and seasonality of 281 individual whales were examined. In all, 60 whales were resighted on different days of which 44 were between different calendar years. The most resightings for one individual was 11 times, seen in six different years, and the longest interval between first and last sightings was about 18 years. A resighting rate of 15.6% of whales at intervals of a year or more indicates long-term fidelity to the region. Shorter intervals of 1–6 months between sequential sightings in the same year may suggest temporary residency. The TF image collection from WSA was compared to TF collections from four other regions, namely Gabon, Cabinda (Angola), Namibia and the Antarctic Humpback Whale Catalogue (AHWC). Three matches were detected between WSA (in late spring or summer) and Gabon (in winter), confirming direct movement between these regions. The capture–recapture data of four different identification features (TF, RDF, LDF and MS) from six successive subsets of data from periods with the highest collection effort (2001–2007) were used to calculate the number of whales that utilise the region, using both closed- and open-population models. Dorsal fins have never been used to estimate abundance for humpback whales, so the different identification features were evaluated for potential biases. This revealed 9–14% incidence of missed matches (false negatives) when using dorsal fins that would result in an overestimate, whereas variation in individual fluke-up behaviour may lower estimates by as much as 57–66% due to heterogeneity of individual capture probability. Taking into consideration the small dataset and low number of recaptures, the most consistent and precise results were obtained from a fully time-dependent version of the Jolly-Seber open-population model, with annual survival fixed at 0.96, using the MS dataset. This suggests that the WSA feeding assemblage during the months of spring and summer (September–March) of the study period numbered about 500 animals. The relationship of these whales to those (perhaps strictly migratory) that may occur in other seasons of the year, and their links to possible migratory routes and other feeding or breeding areas, remain uncertain.

Keywords: abundance, Breeding Stock B, capture heterogeneity, capture–recapture, Chapman’s modified Petersen estimate, Megaptera novaeangliae, migration, photo-identification, Program MARK, site fidelity
Introduction

The west coast of South Africa should function as a near-shore migration corridor for humpback whales Megaptera novaeangliae based on its mid-latitude geographical position and occurrence of such behaviour along the east coast of South Africa (Findlay and Best 1996, Findlay et al. 2011) and at similar locations elsewhere in the Southern Hemisphere (Dawbin 1966, Bryden 1985). However, in the vicinity of Saldanha Bay (at about 33° S), historic and more contemporary observations have shown humpback whales to display seasonal residency from October to February (Olsen 1914, Best et al. 1995, Findlay and Best 1995). Recently, a shore-based survey there with near-complete seasonal coverage (Barendse et al. 2010) has shown that the high relative abundances recorded during these spring and summer months did not correspond to the timing of expected migration peaks, but rather to aggregations of whales feeding on euphausiids (Euphausia lucens) and other crustacean prey.

Humpback whales found in the south-eastern Atlantic are designated to the International Whaling Commission’s (IWC) Breeding Stock B (BSB) (IWC 1998) as included in the ‘Comprehensive Assessment’ of the IWC Scientific Committee (IWC 2010) for Southern Hemisphere populations. This region, particularly the west coast of Africa south of the equator, was characterised by extremely high whale catches from 1908 to 1914 and episodic catches thereafter (Best 1994). The whales from BSB are thought to migrate primarily to Antarctic Areas II (60° W–0°) and III (0°–70° E) for the austral summer, especially to the so-called ‘nucleus feeding area’ located between 10° W and 10° E (Figure 1a, IWC 2010). Based on mitochondrial and more recently nuclear genetic evidence of population substructuring (Pomilla et al. 2005, Pomilla and Rosenbaum 2006, Rosenbaum et al. 2009, Carvalho et al. 2010), BSB has been divided into two breeding substocks, B1 and B2, with the Walvis Ridge or Angola–Benguela Front at about 18° S proposed as a possible boundary (IWC 2010). However, the majority of sampling to date has been limited to only two widely separated localities: on the breeding ground off Gabon (Collins et al. 2008), which is thought to represent BSB1, and off the west coast of South Africa (WSA), which presumably belongs to BSB2. No breeding behaviour has been observed (or is expected to take place) in WSA, so the actual geographical location of the breeding ground for BSB2 remains unknown, and the proposed northern boundary at 18° S would be inconsistent with the sea surface temperature (SST) regimes found for other humpback whale breeding grounds (Rasmussen et al. 2007). The detection of 10 whales biopsied off both Gabon and WSA (Carvalho et al. 2010) using microsatellite genotyping (Palsbøll et al. 1997) has raised questions about the BSB subdivision. Given that the whole coastal region between about 7° and 30° S, comprising the territorial waters of Angola and Namibia (Figure 1a), is mainly unsampled, it remains difficult to construct a conclusive population structure model for the region.

The shore-based observations presented in Barendse et al. (2010) do not add to current understanding of how these humpback whales relate to others in the region as derived from the genetic structure and microsatellite matches between Gabon and WSA (see above). Nor do they provide information on whether the same individuals appear off Saldanha Bay (Figure 1c) during any of the same seasons in different years, or an accurate measure of how many whales utilise the area as a feeding ground. Individual identification through photo-identification (Katona and Whitehead 1981) may help to address these questions. Humpback whales are individually recognisable from two physical features that may be readily photographed: (1) their tail flukes, which includes the trailing edge, and the occurrence of natural marks, scarring, and pigmentation of their ventral surfaces (Katona and Whitehead 1981, Mizroch et al. 1990); and (2) the lateral view of their dorsal fins, which takes into account the shape of the fin, the prominence and distribution of knuckles on the caudal peduncle, and any scarring or pigmentation on the fin and/or flank (Kaufman et al. 1987). Although the use of dorsal fins and lateral body markings has yielded successful matches (Gill et al. 1995), the more distinctive flukes are favoured for use in regional photo-identification catalogues. Such catalogues have been employed widely to identify migratory links (e.g. Stevick et al. 2004), examine regional movement patterns and population structure (e.g. Calambokidis et al. 2001), and calculate population sizes (e.g. Straley et al. 2009).

We present here results from the most comprehensive photo-identification and genetic collection to date from the west South Africa region in order to examine within- and between-year attendance patterns. We investigate inter-regional movements between WSA, Namibia, Gabon, and Antarctic Areas II and III by comparing all available tail fluke collections from these areas. Furthermore, although not specifically collected for this purpose, the type of capture-recapture data obtained from the within-region photographic and genotypic matching may be suitable for the calculation of abundance estimates (Hammond 1986, Hammond et al. 1990). We attempt to estimate the number of humpback whales that may feed in the area during spring and summer, using different approaches including capture-recapture methods on selected subsets of data using different identification features (tail flukes, right and left dorsal fins, and microsatellites). Both closed- and open-population models were applied, as is the norm in many published abundance estimates for large whales, including humpbacks (e.g. Calambokidis and Barlow 2004, Larsen and Hammond 2004, Straley et al. 2009). To our knowledge, this is the first time dorsal fins have been used to calculate abundance for humpback whales, in addition to the more favoured tail flukes. The exposure of fluke, however, can vary for individual whales, which may affect individual capture probability (Perkins et al. 1984, 1985), whereas dorsal fins are always exposed and more easily photographed (Gill et al. 1995). Therefore, we examine potential sources of capture heterogeneity, sampling bias, and error that may result from the use of dorsal fins vs tail flukes as photographic identification features, using double-marked animals (i.e. identified by more than one feature). The results are compared and discussed in terms of the estimation method or model applied, and the identification feature used.
Figure 1: (a) The South-East Atlantic, South-West Indian and Southern oceans showing bathymetry (to 4 000 m), areas of relevance to Breeding Stock B (BSB) Southern Hemisphere humpback whales, the speculated locations of substocks B1/B2, Antarctic Feeding Areas II/III, and suggested nucleus feeding area for BSB whales (10° W–10° E, shown by dashed grey lines), and collection areas for regional photo-identification catalogues; (b) detail of WSA and extent of collection effort from various sources; (c) detail of Saldanha/St Helena Bay area showing the locations where humpback whale data were collected during four major research projects, 1993–2007 (also see Tables 1 and 2).
Material and methods

Data collection and sighting database

The sighting database and photographic catalogue were compiled from a number of data sources (Table 1), but as a minimum requirement for inclusion they had to be collected from within the Exclusive Economic Zone (EEZ) of South Africa, west of Cape Agulhas (20° E). These included data from humpback whales encountered incidentally during research work directed at other cetacean species, or during routine multidisciplinary scientific cruises in the region, over the period 1983–2008 (Figure 1b). It further included all boat study dedicated to humpback whales at Cape Columbine in 1993, described by Best et al. (1995) (Figure 1c). It was attempted throughout to photograph the ventral side of the tail flukes and both left and right sides of the dorsal fin, and from 1993 to collect a biopsy from every whale encountered. However, any whale that had at least one of these features recorded was included, and the date (day, month and year) and locality (latitude and longitude) of the sighting noted. In most cases, additional data (including group size, composition and behaviour, SST, depth, and duration of encounter) were also collected. Discrimination between individuals in the field (and association of specific images/biopsy attempts with individuals) was aided by onboard notes and sketches of body features, and by recording all photographic (film roll/ data card numbers and frames) and biopsy sampling effort for each individual. This information was later used in the database to associate identification features with specific individuals seen during a sighting.

Prior to 2004, most images were recorded on high-speed (ISO 400 and higher), black-and-white or colour-negative, and colour-positive film using motor-driven 35 mm single lens reflex (SLR) cameras with 100–300 mm manual focus lens zoom lenses; from January 2005 onwards, these were replaced by digital autofocus SLR cameras. Digital images, and film frames scanned at 600 dots per inch (dpi), were cropped to maximise the coverage of the area of interest (i.e. tail flukes, or dorsal fin plus caudal peduncle), and imported into the sighting database in the JPG file format. Black-and-white negatives were scanned according to an unpublished protocol (Santos-Tieder et al. 2003, cited in IWC 2004). Each image was individually assessed for photographic quality and orientation of the subject, and assigned a score based on a 5-point scale (1 = not useable, 2 = poor, 3 = fair, 4 = good and 5 = excellent). Every tail fluke (TF) image was further classified according to its ventral pigmentation pattern (or ‘type’) on a scale of 1–5, where 1 is all white (no central black bar between the left and right flukes) and 5 all black (see Rosenbaum et al. 1995). Flukes were rated for the part visible above water, i.e. whole, left fluke only, right fluke only, and trailing/leading edge. An additional classification type ‘0’ was introduced for TF where it was impossible to assign types 1–5, either due to the unfavourable orientation or partial obscuring of the subject, or where the tail flukes were severely scarred or mutilated due to injury, such as killer whale Orcinus Orca bites. Images were also assigned a score from 1 to 5 for individual distinctiveness of the subject, although this rating was not used in any of the present analyses.

Skin biopsies were collected using the Paxarms rifle system (Krützen et al. 2002). All biopsy heads were sterilised by flaming after use. Samples were placed into individually labelled cryogenic tubes filled with a NaCl-saturated, 20% dimethylsulphoxide (DMSO) solution and placed on ice bricks in a cooler box. At the end of each day all skin samples were stored in a domestic freezer (−5 °C) until they could be transferred to a −15 °C freezer at the laboratory in Cape Town. Processing of samples was carried out at the Sackler Institute for Comparative Genomics (American Museum of Natural History).

Within-region matching

The matching described below was done separately for each identification feature. Thumbnail (100 dpi) or medium-resolution (200/250 dpi) copies of the original pictures for all useable images (i.e. with photo and orientation quality ratings of poor and better) were viewed on 38–48 cm (15–19 in) thin film transistor (TFT) computer screens. Original (large format) images were viewed for final decision-making. Tail flukes were compared by pigmentation type to reduce the number of possible comparisons, first to all images of the same type and then to all images from the preceding and following types (e.g. type 2 was compared to types 1, 2 and 3). Type 0 flukes were compared to all available images from all other types. In the case of dorsal fins, each image was compared with every other image. Within-year matching was carried out first, i.e. checking for matches of the same individuals on different days in the same year. Once completed, representative images of individual whales from each year were

Table 1: Photographic and genetic contributions to the west South African humpback whale database from various projects and sources.

<table>
<thead>
<tr>
<th>Project description</th>
<th>Study years</th>
<th>Number of images/biopsies collected**</th>
<th>Individuals identified</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td>Total TF RDF LDF Biop.</td>
<td></td>
</tr>
<tr>
<td>Miscellaneous contributions</td>
<td>1983–2007</td>
<td>143 96 30 17 1</td>
<td>32</td>
</tr>
<tr>
<td>Cape Columbine humpback *</td>
<td>1993</td>
<td>104 30 37 37 6</td>
<td>9</td>
</tr>
<tr>
<td>Saldanha Bay humpback whale*</td>
<td>2001–2003</td>
<td>739 173 294 272 104</td>
<td>135</td>
</tr>
<tr>
<td>Saldanha Bay/St Helena Bay southern right whale*</td>
<td>2003–2007</td>
<td>736 192 300 244 92</td>
<td>95</td>
</tr>
<tr>
<td>Entire database</td>
<td>1 820</td>
<td>510 694 616 216</td>
<td>289</td>
</tr>
</tbody>
</table>

* Indicates projects by the Mammal Research Institute
** Numbers include all images and biopsies collected and incorporated into the database. It does not take photo quality or matches into consideration TF = tail fluke; RDF = right dorsal fin; LDF = left dorsal fin
compared in chronological order to those of the subsequent year in the database and matches identified. The processes of within- and between-year matching were repeated by a second person. Where a match disagreed, it was reviewed and a consensus decision made to accept or reject it. Once all matching was completed, the best image(s) available per individual and identification feature were selected for representation in the overall catalogue, and a unique identification number assigned per identification feature.

The methodology for genotyping using 10 microsatellite loci is detailed in Carvalho et al. (2010). Each biopsy was associated to an individual sighting incident by its original biopsy number. In the case of a positive match between two skin biopsies, the laboratory code assigned to the earliest collected sample was retained as the identification number for that individual.

**Periodicity and seasonality of resightings**

Although matching was carried out for each feature independently, a maximum of four identification features, viz. tail flukes (TF), right dorsal fins (RDF), left dorsal fins (LDF) and microsatellite (MS), could be collected for an individual whale at any given encounter. Wherever a common identification feature was identified between two or more different sightings, these could be linked. Thus, a full sighting history could be built based on all matches made through all available identification features between different encounters, even though these were not all collected at every sighting. It is important to note that failure to positively link one feature to another for the same individual could result in missed matches between different sightings. The problems of having multiple separate records for the same animal in a combined catalogue feature was warranted in order to optimise the sample size especially when dealing with large numbers of individuals. However, given the small total number of humpback whales identified, we believe the use of combined identification features was warranted in order to optimise the sample size for the purposes of examining trends in the growth of the catalogue and attendance patterns.

Within- and between- (calendar) year occurrences of resighted individual whales were examined using combined identification features (genotype and photos of usable quality) for the entire database. The time interval between the dates of first and last sightings (excluding the first day) was calculated for all individual whales that were resighted on different days, both within and between years. For whales sighted on successive days, the time between sightings was assumed to be one day, i.e. rounded up to 24 h. Between-year time calculations took leap years into account. The number of days between sequential sighting events was also calculated for each individual whale. The seasonality of resightings for the entire sighting database was examined by sorting them by month, and separated on the basis of their overall resighting histories, i.e. seen only once, resighted within years only, and resighted between different calendar years. Note that the latter may have included some within-year sightings, but were not included in the ‘within-year only’ category.

**Between-region photographic matching**

The representative images of 154 individual humpback whales identified by TF that resulted from the WSA within-region matching (see above) were compared to TF collections from four other regions (see Figure 1a for localities): Cabinda — In all, 25 individual whales of which identification pictures of TF (45 images in total) were taken during September 1998 off Cabinda, Angola, around oil production platforms some 50 nautical miles south of Congo River mouth (Best et al. 1999) were compared to the WSA, Namibia and Gabon catalogues; Gabon — A total of 1 297 individuals, represented by 9 776 images collected between 2001 and 2006, were compared to the WSA and Cabinda images. The database, area of collection, and matching procedures are fully described by Collins et al. (2008); AHWC (feeding Areas II/III) — The Antarctic Humpback Whale Catalogue (AHWC) is a compilation of almost 5 000 photographs (TF, LDF and RDF) taken by miscellaneous contributors, both by scientists and non-scientists since 1987. The images originate from regions throughout the Southern Hemisphere, and the overall aim of the AHWC is to investigate movements of humpback whales between the Southern Ocean and lower latitude waters through an internationally collaborative project (Allen et al. 2008). It is currently maintained by the College of the Atlantic (Maine, USA) and is available on the web-based photo-sharing platform Flickr® (http://www.flickr.com/ahwc). The photostream can be viewed as a whole, or by sets, using the search tool to select any combination of tags or text, such as TF pigment type or locality of picture (for example, the tag ‘T’1 areaall’ would display all images of type 1 from Area III) (J Allen pers. comm.). The type 0 is not used in the AHWC. A total of 186 images representing 130 individuals, tagged as being from Areas II and III, were compared to the WSA images.

Namibia — There is presently no formal humpback whale catalogue for Namibia, but images have been collected at Walvis Bay (23°00′ S, 14°30′ E) during research cruises directed at Heaviside’s *Cephalorhynchus heavisidii* and bottlenose *Tursiops truncatus* dolphins, or by dolphin- and whale-watching operators in winter (June–August) and summer (January–March) of the years 2008, 2009 and 2010. Preliminary sorting and matching of these yielded 35 individuals (61 images).

Images of both whole and partial TF of all quality ratings except ‘not useable’ were considered. The AHWC does not catalogue non-useable images as individuals (J Allen, College of the Atlantic, Maine, pers. comm.). No matching was conducted between the Gabon catalogue and the images from the AHWC and Namibia. Representative images of each individual in one database were systematically compared to those of the other, bracketed by fluke type (as described above for within-region matching) to avoid mismatches due to the variable assignment of TF types. All matches were checked and confirmed by a second person.

**Abundance estimates**

**Catalogue size adjusted for annual survival**

For each of the four identification features, a measure of the absolute minimum abundance was derived from the number of individual whales contained in the respective databases. This was done similarly to the method used by Straley et al. (2009), where the number of whales ($\hat{N}$) alive in any given
year (\(\chi\)) is calculated by adding the number of unknown (or
‘new’) individuals identified in that year (\(\hat{n}_j\)) to the number
estimated to have survived from the preceding year (\(\hat{N}_{j-1}\)),
the latter being adjusted by an annual survival rate (\(\phi\))
(Equation 1). The term \(\hat{N}_{j-1}\) is the sum of \(\hat{n}_j\) and \(\hat{N}_{j-2}\) (again
adjusted with \(\phi\)), and so forth. No variance can be calculated.

\[
\hat{N}_j = \hat{n}_j + \phi (\hat{N}_{j-1})
\]  

(1)

The value for \(\phi\) was set at 0.96 as calculated for humpback
whales in the North Pacific ( Mizroch et al. 2004). Although this value is probably lower for non-adults (Zerbini et al.
2010), it is considered a reasonable estimate for annual adult
survival, given that the area is not a breeding ground and very
few calves were seen (Barendse et al. 2010).

Data selection for capture–recapture estimates
The only time period for which sufficient data were available for
several years in sequence, and offered adequate seasonal coverage to permit estimation of abundance for
whales that engage in spring/summer feeding, occurred during 2001–2007 (Table 2). This included the sighting data from the boat-based component of the work described in
Barendse et al. (2010) (see above), as well as humpback whales encountered during work on feeding southern right
whales Eubalaena australis (2003–2007) at Saldanha Bay
(in September) and St Helena Bay (in October–December,
and rarely in January) — note that this study had no shore-
based watch (see Table 1). By restricting the data subsets to only certain seasons, the possible heterogeneity in
capture probability introduced by different seasonal attendance patterns of individuals should be reduced. Six successive capture occasions \((j)\) of six months each were identified,
starting in September of one year and ending in February the
following year (e.g. \(j_1 = 01\) September 2001 to 28 February
2002, both dates inclusive) (see Appendix).

Variation in photographic quality and the distinctiveness of
natural marks can affect the ability to correctly match different
photographs of the same individual, and hence the likelihood of
a successful resighting (Hammond 1986, Gunnlaugsson and
Sigurjónsson 1988, Friday et al. 2000, Stevick et al.
2001). For example, on images of poor quality, highly distinctive individuals may still be identified while matches of less distinctive animals are more likely to be missed (i.e. an
increased probability of false negatives). To reduce such
errors, the commonly used approach of excluding images
below a certain quality was applied (e.g. Cerchio 1998, Friday et al. 2008, Straley et al. 2009). In this case, those of quality
and/or orientation rating of ‘poor’ and ‘not useable’ were not
used for capture–recapture calculations, and no partial TF
pictures (halves or trailing edges) were included.

Closed-population model
The two-sample Chapman’s modified Petersen (CMP)
estimator (Seber 1982) has been used elsewhere to calculate the size of feeding aggregations of humpback whales (e.g.
applied over relatively short time periods (e.g. one-year
intervals), this is considered an acceptable approach for
a long-lived mammal with relatively low rates of natural
mortality and recruitment, despite such populations generally
not meeting the assumptions of closed-population models. These assumptions (adapted from Seber 1982), applicable
when using natural marks, are: (1) a constant population
during the sampling period (no immigration or emigration,
or births/deaths); (2) no loss of marks between sampling
periods; (3) all marks are correctly recorded; (4) all whales
have an equal chance of being recorded in the first sample;
and (5) both previously identified and newly sighted whales have equal probability of recapture in subsequent samples.
We employed the CMP estimator here because of its
relative simplicity, and to illustrate issues that relate to the
different identification features used (see later), using the
formula (Seber 1982):

\[
N^* = \frac{(n_1 + 1)(n_2 + 1)}{(m_2 + 1)} - 1
\]  

(2)

where \(N^*\) is the estimated population size, \(n_1\) the number of
whales identified during \(j_1\), \(n_2\) the number of whales identified
during \(j_2\), and \(m_2\) the number of whales identified (i.e.
matched) in both periods. The estimated variance \((\nu \text{ or } \sqrt{\text{var}})\) of \(N^*\) and the estimated coefficient of variation \((CV^*)\) of \(N^*\)
were calculated according to Seber (1982):

\[
\text{var}(N^*) = \frac{(n_1 + 1)(n_2 + 1)(n_1 - m_2)(n_2 - m_2)}{(m_2 + 1)^2(m_2 + 2)}
\]  

(3)

and

\[
CV^* = \sqrt{\text{var}(N^*)}/N^*
\]  

(4)

Confidence intervals (95%) for the CMP estimator were
calculated with the log-normal transformed method as proposed by Burnham et al. (1987):

\[
r = \exp\left\{1.96\sqrt{\ln(1+(CV^*)^2)}\right\}
\]  

(5)

The lower confidence interval (CI) was calculated by dividing
\(N^*\) by \(r\), and the upper CI by the product of \(N^*\) and \(r\).

The CMP calculation was restricted to the first pair of
capture periods \((j_1–j_2)\) as these were the only ones with the
primary effort directed at humpback whales, with the
largest sample sizes, and where recaptures were detected
for all identification features. Furthermore, sampling during
\(j_1–j_2\) occurred at the same site of limited extent (i.e. within
\(\pm 25\) km radius from North Head, Saldanha Bay); this should
reduce capture heterogeneity, a factor not accounted for
by the CMP estimator between individuals, or over time
(Hammond 1986). Such heterogeneity is regarded as highly
likely to be a factor for all natural populations, resulting in
underestimation of the true size of the population, which
sometimes can be considerable (Carothers 1973).

Open-population models
Maximum-likelihood models of the Jolly-Seber (JS) type
(Jolly 1965, Seber 1965, Schwarz and Seber 1999) are
Table 2: Annual collection effort of photo-identification and genetic data that contribute to the west South African humpback whale database, expressed as number of days on which at least one identification image or biopsy was collected or ‘collection days’. Numbers in brackets indicate total days on which boat was deployed, when known; ‘x’ indicates months with no boat effort during dedicated Mammal Research Institute studies. Months within dashed outline indicate West Coast Heaviside’s dolphin study period; light-grey shading indicates dedicated humpback whale study at Saldanha Bay (with shore-based observations); dark-grey shading indicates boat-based study on southern right whales at St Helena Bay. Bold numbers in 2001–2007 show those months used for abundance estimates

<table>
<thead>
<tr>
<th></th>
<th></th>
<th></th>
<th></th>
<th></th>
<th></th>
<th></th>
<th></th>
<th></th>
<th></th>
<th></th>
<th></th>
<th></th>
<th></th>
</tr>
</thead>
<tbody>
<tr>
<td>1983</td>
<td>2</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td>2</td>
</tr>
<tr>
<td>1984</td>
<td></td>
<td>1</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td>1</td>
</tr>
<tr>
<td>1988</td>
<td>1</td>
<td></td>
<td></td>
<td></td>
<td>1</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td>2</td>
</tr>
<tr>
<td>1989</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td>1</td>
</tr>
<tr>
<td>1990</td>
<td>1</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td>1</td>
</tr>
<tr>
<td>1992</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td>1</td>
</tr>
<tr>
<td>1993</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td>1</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td>7 (18)</td>
</tr>
<tr>
<td>1997</td>
<td>1</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td>2</td>
</tr>
<tr>
<td>1999</td>
<td>X</td>
<td>3 (13)</td>
<td>1 (13)</td>
<td>X</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td>4 (26)</td>
</tr>
<tr>
<td>2000</td>
<td>0 (4)</td>
<td>4 (13)</td>
<td>1 (16)</td>
<td>0 (6)</td>
<td>X</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td>5 (39)</td>
</tr>
<tr>
<td>2001</td>
<td>0 (8)</td>
<td>0 (14)</td>
<td>1 (15)</td>
<td>1 (7)</td>
<td>X</td>
<td>X</td>
<td>1 (7)</td>
<td>1 (14)</td>
<td>4 (8)</td>
<td>5 (11)</td>
<td>3 (10)</td>
<td>5 (14)</td>
<td>5 (9)</td>
</tr>
<tr>
<td>2002</td>
<td>2 (9)</td>
<td>2 (2)</td>
<td>X</td>
<td>X</td>
<td>X</td>
<td>X</td>
<td>X</td>
<td>X</td>
<td>1 (2)</td>
<td>3 (11)</td>
<td>3 (12)</td>
<td>0 (5)</td>
<td>16 (41)</td>
</tr>
<tr>
<td>2003</td>
<td>3 (9)</td>
<td>X</td>
<td>X</td>
<td>X</td>
<td>X</td>
<td>X</td>
<td>X</td>
<td>X</td>
<td>2 (8)</td>
<td>5 (15)</td>
<td>4 (9)</td>
<td>3 (10)</td>
<td>17 (51)</td>
</tr>
<tr>
<td>2004</td>
<td>5 (2)</td>
<td>1</td>
<td>X</td>
<td>X</td>
<td>X</td>
<td>X</td>
<td>X</td>
<td>X</td>
<td>2 (9)</td>
<td>4 (18)</td>
<td>3 (18)</td>
<td>x</td>
<td>12 (51)</td>
</tr>
<tr>
<td>2006</td>
<td>X</td>
<td>X</td>
<td>X</td>
<td>X</td>
<td>X</td>
<td>X</td>
<td>X</td>
<td>X</td>
<td>0 (1)</td>
<td>1 (16)</td>
<td>6 (17)</td>
<td>3 (7)</td>
<td>12 (41)</td>
</tr>
<tr>
<td>2007</td>
<td>0 (2)</td>
<td>0 (7)</td>
<td>X</td>
<td>X</td>
<td>X</td>
<td>X</td>
<td>X</td>
<td>X</td>
<td>0 (1)</td>
<td>2 (8)</td>
<td>0 (8)</td>
<td>2 (9)</td>
<td>1</td>
</tr>
<tr>
<td>All</td>
<td>14</td>
<td>14</td>
<td>4</td>
<td>2</td>
<td>1</td>
<td>5</td>
<td>13</td>
<td>28</td>
<td>30</td>
<td>12</td>
<td>135</td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

frequently used when the assumption of population closure is unlikely to be met, and when data from multiple capture periods are available. The POPAN option, included in the software Program MARK 5.1 (White and Burnham 1999, Schwarz and Arnason 2006), is one of the JS model formulations most readily available to biologists (Arnason and Schwarz 1999). It has therefore enjoyed wide application for generating population estimates from photographic and genotypic capture–recapture data for several cetacean and other large marine species, including humpback whales (Larsen and Hammond 2004), Indo-Pacific bottlenose dolphins Tursiops aduncus (Reisinger and Karczmarski 2010), killer whales (Reisinger et al. 2011), North Pacific right whales Eubalaena japonica (Wade et al. 2011), and whale sharks Rhincodon typus (Meekan et al. 2006). The latter two examples involved a very small population, and one for which limited data were available, respectively.

The POPAN model estimates the following parameters:

- the super-population size \( N \);
- the apparent survival rate \( \phi \); the probability of entry into the population, or ‘\( P_{\text{en}} \), with the alternative notations of \( b \) or \( \beta \) (the latter is used here); and capture probability \( p \) at capture occasion \( j \) (Schwarz and Arnason 2006). The prescribed link function (GC White, Program MARK Help files), namely the Logit link for \( \phi \) and \( p \), and multinomial Logit (MLogit) link for \( \beta \), were used.

Different variations of the model were applied to datasets for six successive capture occasions \( (j-j_0) \) for all four identification features (TF, LDF, RDF and MS) including all parameters fixed (.), full time-dependence (t) for \( \phi \), \( \beta \) and \( p \), and with \( \beta \) fixed at the biologically realistic value of 0.96 (see above). While the \( \beta \) parameter accounts for the contribution of births to the overall entry rate (Arnason and Schwarz 1999), and although there are published annual rates of increase (ROI) available for humpback whales (see Zerbini et al. 2010), no attempt was made to fix this at a specific value, given that our data are not likely to be (fully) representative of a discrete breeding population. Selection of the best models was done using the quasi-Akaike’s information criterion (QAICc), adjusted for small sample sizes as implemented in MARK (Cooch and White 2006).

Biases in abundance estimates derived from different photographic identification features

Given that dorsal fins have never been used to calculate abundances for humpback whales, their reliability as a naturally marked feature for this purpose is untested. It is expected that identification features with less information or that are less distinctive would be more difficult to match, which can result in misidentification (Hammond 1986), as is the case for other species where dorsal fins are used (Gowans and Whitehead 2001). Therefore, we examined the incidence and effect of missed matches when using dorsal fins. Furthermore, we assessed the possible impact of variation in individual fluking behaviour (on estimates) as it is a known idiosyncratic behavioural feature (see Perkins et al. 1984, 1985). Also, there was a sense during the data collection that it was more difficult to photograph the flukes of some individuals, a notion reinforced by fewer individuals identified by this feature compared to dorsal fins (see below). While we acknowledge that the use of genotypes is not completely free from error and may cause an upward bias in abundance estimates on account of misidentification of microsatellites (see Lukacs and Burnham 2005, Wright et al. 2009), detailed consideration of this issue is beyond the scope of this paper. However, we did compensate for it where applicable or possible in the analyses below.

Downloaded by [University of Pretoria] at 01:36 19 January 2012
Tests for false negative rates
Microsatellites were used as an independent (non-photographic) identification feature and all individuals \((n = 32)\) that were identified by this feature and resighted on different days were used as the sample. For each capture occasion (day), it was assessed whether a specific photographic feature of useable quality (>poor) was recorded; then, whether or not a specific feature confirmed the matches made by microsatellite. The sample size per identification feature was the number of times both a MS match and a photograph of the feature in question were available (‘matching opportunities’). Failure to detect a photographic match constituted a false negative. As a simple test to quantify the positive bias caused by the detected false-negative error rate \((e)\), the pairwise CMP estimator (see Equation 2) was calculated for the applicable dataset, using the false-negative correction developed by Stevick et al. (2001). The identification events \((s)\) per sampling period \((j)\) were taken as the sum of every time a whale was identified as an individual, excluding same-day resightings, therefore assuming that the boat crew recognised such individuals in different groups on the same day. Thus, to correct for the higher-than-actual total number of whales ‘identified’ due to missed matches within each sampling period, the numbers of individuals identified during \(j_1\) and \(j_2\) \((n_1\) and \(n_2\)) were calculated as:

\[
\begin{align*}
    n_j^* &= \frac{n_j - e \cdot s_j}{1 - e} \\
    n_j^2 &= \frac{m_2}{1 - e}
\end{align*}
\]

The number of individuals matched between these samples \((m_j)\) was increased by the error factors to correct for missed matches between \(j_1\) and \(j_2\) in the following manner:

\[
m_j^2 = \frac{m_j^2}{1 - e}
\]

A comparison of the resultant population estimates with the uncorrected estimates provided estimates of the magnitude \((\%\) of overestimation.

Variation in recording of tail flukes for resighted whales relative to other features
All whales resighted on different days \((n = 60)\) were used as the sample, and the identification features collected during intercepts on these different days were compared. First, the number of times TF were recorded (of any photographic quality) during all intercepts of resighted whales was compared to that of other features. Second, the frequency with which TF were recorded in the case of multiple resightings was examined. Third, the duration of intercepts where TF were recorded was compared to those where no TF were recorded. Finally, the probability of recording TF or dorsal fins (left or right) for an individual whale was calculated by counting the number of intercepts during which the feature was recorded, and expressing it as a fraction of the total number of times that the resighted whale was intercepted.

Use of double marks
We used TF as one type of mark, and LDF, RDF and MS respectively as alternative marks. For the two adjacent sampling periods \((j_1\) and \(j_2\)), the \(n_j\) consisted of animals that were identified by both TF and the other mark in question, i.e. double-marked animals. The \(n_j\) consisted of the total number of whales identified by either TF, or the alternative mark in the following sampling period, with recaptures \((m_j)\) being those double-marked animals that were identified by whatever feature was used for \(n_j\). This approach is intended to compare the relative capture probabilities of the two marks used: if they are equal, then recapture rates (and by inference, abundance estimates) should be similar whichever feature is used for the second sample. During the calculation using the CMP estimator (Equation 2), an error correction factor \((e)\) was applied to dorsal fins and MS similar to that described above (i.e. \(n_j\) was adjusted downward [Equation 6] and \(m_j^2\) adjusted upward [Equation 7]). However, \(n_j\) was left unadjusted because the animals were already identified without error from their TF. The correction factors used for dorsal fins were those calculated from LDF and RDF false-negative tests (see below). When MS was used as the alternative identification feature, it was adjusted by the mean allelic error rate of 0.065, calculated for the samples collected off WSA (IC unpublished data).

Results

Range and seasonality of collection effort
On account of the \(ad hoc\) and variable manner in which much of the photographic and genetic data were obtained, effort is loosely defined here as ‘collection days’, i.e. any day on which such data were collected. There were only 28 such days from 1983 to 2000, compared to 106 over the next eight years (Table 2). The greatest number (and days with boat availability) of collection days occurred between 2001 and 2006 during the two studies at Saldanha Bay and St Helena Bay (reported earlier) and made the greatest overall contribution in terms of number of images and individuals identified after matching was completed (Table 1). Other notable periods of data collection were during the earlier study at Cape Columbine (Best et al. 1995) and incidental humpback sightings made during a project on Heaviside’s dolphins (described in Elwen et al. 2009). Collection days, as a proportion of days where a boat was deployed, ranged from 12.8% (in 2000) to a maximum of 38.8% in 1993, and most years ranged around 20–30% (Table 2). Overall, at least one collection day was recorded during any given month, but effort was not evenly distributed across seasons. The autumn and winter months (March–August) had the poorest overall coverage with 10 or less collection days per month, whereas spring and summer months (September–February) were better sampled. Most collection days occurred in November \((n = 30)\) and fewest in June \((n = 1)\) (Table 2).

The spatial extent of miscellaneous data collection along the West Coast was fairly extensive (approx. 700 km between the northern- and southernmost sites; Figure 1b). However, the majority of data were collected within a fairly limited area of about 1 \times\ 1 degree latitude/longitude grid square, no farther than 25 km from the shore (Figure 1c), and included the major study sites mentioned above.

Within-region matching

Sighting database/catalogue
The WSA catalogue up to February 2008 included a total of 1 820 images, made up of 510 TF, 694 RDF and 616
LDF (Table 1), representing 446 individual sighting histories collected during 225 boat intercepts/encounters. Excluding images that were deemed not useable, 154 individuals were identified using only TF, 237 by RDF and 230 by LDF (see Table 4). Microsatellite genotyping of 216 skin biopsies yielded 56 samples matched to one or more other samples, representing 156 individuals, three of which were identified by microsatellite only (i.e. were not photographed). By linking different individual identification features to common sightings, a total of 289 individual whales was identified with ‘combined features’, although images of eight animals were not useable and were thus excluded \((n = 281)\). Few animals \(<10\) per annum \) were identified before the advent of dedicated field work in 2001 (Figure 2), when most individuals were identified in a single year \((n = 80)\). New additions remained at fairly high levels for the following five years \((>25\) individuals per annum\), although there was a steady decrease in the growth rate of the database (Figure 2).

**Resighting rates, intervals and seasonality**

Using combined identification features \((n = 281)\), 214 individual whales were seen once only, seven were resighted on the same day (i.e. in more than one group), and 60 \((21.35\%)\) on different days. Some 44 whales were resighted between calendar years, the majority only once \((n = 30)\), followed by twice \((n = 7)\) to a maximum of five resightings (i.e. in six different years). Only 12 of these between-year sightings were not seen on multiple occasions in the same year, with one individual recorded a total of 11 times (the same whale that was seen in six different years).

The shortest interval between first and last sighting events was one day and the longest 18 years, with the mean interval being 3.4 years and the median 1.5 years. Most whales were resighted within one year \((n = 23)\), followed by a 1–2 year interval \((n = 17)\). For 14 whales, the interval was longer than four years, and for six of these, longer than 12 years (Figure 3). A breakdown of the time intervals between sequential sightings (Figure 4) of all resighted whales showed that most individuals were resighted on the same day \((35\) times), or within a week of the previous sighting. Resightings at intervals of more than a week, but less than six months, were relatively few \(<10\). The next most commonly observed resighting intervals were at 6–12 months and 1–2 years (Figure 4). Intervals of between 2 and 3 years, and longer than 5 years, were recorded less than 10 times each, whereas intervals between 3 and 5 years were uncommon.

None of the 32 individual whales seen during winter (June–August) was resighted (Figure 5). During all other months some of the whales seen were resighted on other occasions, the majority between calendar years. Between October and January, a small proportion of resighted individuals were same-year resightings only. However, from February to May all resighted individuals were between years, and 50% or more of whales seen during these three months had been seen previously (Figure 5).

**Between-region matching**

None of the images from Cabinda or Namibia matched a whale in any of the catalogues they were compared with. Three matches were made between the WSA and Gabon catalogues, and two between the WSA catalogue and the Area II/III images contained in the AHWC (Table 3; also see Figure 1b). Three of these whales (TF-ZAW-01-005, TF-ZAW-03-017 and TF-ZAW-04-005) were also resighted in different years off WSA (Table 3). The matches with the AHWC were found to be with two humpbacks sighted together on the first day of the IWC-SOWER (Southern Ocean Whale and Ecosystem Research) cruise that departed on 22 December 2005 from Cape Town for the Antarctic; the images were inaccurately tagged in the database as being from Area III. Both were males (determined from biopsies collected off WSA) and one animal (TF-ZAW-05-007) was seen less than a month before in St Helena Bay, some 150 km to the north (Table 3). The second animal had been seen previously in St Helena Bay.
in December 2004 when it was accompanying a cow–calf pair, and was identified as a possible yearling calf. It was also seen subsequently, on 22 November 2006 (also in St Helena Bay) with a different female, when several defecations were observed, presumably an indication of recent feeding.

A northward transit with duration of about 230 days between sequential sightings was recorded for two of the WSA-Gabon matches (Table 3). Shorter southward transit periods (40–80 days) between Gabon and WSA for sightings in the same calendar years were recorded. One male (TF-ZAW-01-005) provides an interesting perspective in that it was sighted off WSA during the years before and after being photographed in Gabonese waters. It was first seen off Saldanha on 16 December 2001 as part of a group that defecated. On 6 August 2002 (233 days later) it was identified off Gabon, before appearing off Saldanha Bay 88 days later, where it apparently remained in the vicinity for a period of over two months, to be resighted on 14 January 2003. It was again resighted on 7 November 2006 when it approached the research boat during a plankton haul in St Helena Bay (Table 3).

**Abundance estimates**

Quality control criteria excluded 122 out of 1,409 images (all photographic features) from the datasets used for abundance estimates. The summary capture–recapture tables of data used in the models (see Appendix) show overall small

![Figure 3: Intervals (in weeks and years) between the first and last sighting events for 60 humpback whales resighted on different days off west South Africa](image)

![Figure 4: Frequency of occurrence of time interval between successive sightings of 67 individually identified humpback whales off west South Africa (n = 157 sighting records)](image)
sample sizes and few recaptures, especially for TF. During the first two sampling periods, more individuals were identified by dorsal fins than other methods, and more matches were made, although for the remaining four periods most whales were identified by MS. The latter feature generally had the highest recapture rate relative to the total number of whales identified, and between all pairs of recapture periods. Fewer resightings were recorded for LDF than RDF.

**Adjusted catalogue size**
The total number of individual humpback whales in the database (after correction for annual survival) represented by TF was over 30% lower than for LDF or RDF, whereas it was almost the same as MS, bearing in mind that biopsy sampling only started in 1993 (Table 4). Given that failure to match dorsal fins (or genotypes) that belong to the same individual contained in the database would inflate the catalogue size for the relevant feature, the total catalogue size was reduced (after correction of survival) by the calculated false-negative rate for the respective feature (see Table 7). The numbers of whales identified by LDF and RDF were still greater than for TF (by 22.4% and 29.7% respectively).

**Closed-population model**
The CMP estimates from TF data were the lowest overall, even less than the lower 95% confidence intervals of the

---

**Table 3:** Details of sightings (date and position) of humpback whales involved in photographic matches between tail fluke catalogues of WSA (‘ZAW’) and Gabon (‘Gab’), and WSA and AHWC (sex determined from biopsies), and time elapsed between sequential resightings.

<table>
<thead>
<tr>
<th>WSA ID (sex)</th>
<th>Date (and position) photographed off west South Africa</th>
<th>Matched to other catalogue ID</th>
<th>Date (and position) photographed in other region</th>
<th>Time between consecutive between-catalogue resightings (and direction of movement)</th>
</tr>
</thead>
<tbody>
<tr>
<td>TF-ZAW-03-017 (F)</td>
<td>2003/01/13 (33.013° S, 17.774° E) and 33.064° S, 17.825° E; 2003/01/14 (32.702° S, 17.99° E) 2008/02/05 (33.03° S, 17.875° E)</td>
<td>TF-Gab-03-124 2003/09/04 (9.264° S, 1.928° E)</td>
<td>234 days (N); 1615 days (S)</td>
<td></td>
</tr>
<tr>
<td>TF-ZAW-04-005 (F)</td>
<td>2004/11/08 (32.665° S, 17.988° E)</td>
<td>TF-Gab-04-045 2004/09/26 (9.264° S, 1.928° E)</td>
<td>43 days (S)</td>
<td></td>
</tr>
<tr>
<td>TF-ZAW-01-005 (M)</td>
<td>2001/11/16 (33.021° S, 17.86° E); 2002/11/02 (33.005° S, 17.849° E); 2003/01/14 (33.031° S, 17.825° E) and 32.674° S, 17.877° E; 2006/11/07 (32.674° S, 17.935° E)</td>
<td>TF-Gab-02-299 2002/08/06 (9.264° S, 1.928° E)</td>
<td>233 days (N); 88 days (S)</td>
<td></td>
</tr>
<tr>
<td>TF-ZAW-05-007 (M)</td>
<td>2005/11/24 (32.551° S, 18.026° E)</td>
<td>ahwc3054 2005/12/22 (33.859° S, 18.278° E)</td>
<td>28 days (S)</td>
<td></td>
</tr>
<tr>
<td>TF-ZAW-06-014 (M)</td>
<td>2004/12/01 (32.703° S, 17.888° E); 2006/11/22 (32.973° S, 17.856° E)</td>
<td>ahwc3055 2005/12/22 (33.859° S, 18.278° E)</td>
<td>386 days (S); 335 days (N)</td>
<td></td>
</tr>
</tbody>
</table>
estimates for all other identification features (Table 5) and less than the adjusted TF catalogue size. Tail fluke estimates were between 70% and 80% lower than uncorrected dorsal fin estimates, and about half of the uncorrected MS estimates. Even when these features were corrected for false-negative errors (see below), the TF estimates were still 45–75% smaller. The highest overall $N^*$ was from RDF, then LDF and MS, although all estimates had fairly wide 95% CIs. The estimates from the genotypic recaptures had the lowest CVs.

**Open-population models**
Model configurations with all or most parameters constant or fixed ($\{\phi, \beta, p\}$ and $\{\phi_{0.96}, \beta, p\}$), or with capture probability set to vary between capture periods, and other parameters constant or fixed ($\{\phi_{0.96}, \beta, p\}$), showed very poor fit, or failed to converge, and were not considered. The remaining model variants applied were:

1. full time-variance for all parameters ($\{\phi, \beta, p\}$);
2. full time-variance for two parameters with $\phi$ fixed at 0.96 ($\{\phi_{0.96}, \beta, p\}$);
3. $\phi$ fixed at 0.96, $\beta$ set to vary fully over time, and $p$ constant ($\{\phi_{0.96}, \beta, p\}$).

Using the $\Delta$AIC$_c$ as indication, model 2 showed the best fit for all identification features, except for the LDF data for which the full time-variant (model 1) fitted best (Table 6).

### Table 4: Minimum photographic and genetic individual catalogue size, by year and identification feature for WSA humpback whales.
Correction for apparent survival ($\phi$) is 0.96 (from Mizroch et al. 2004); correction for matching error ($e$) is calculated false-negative rates for RDF/LDF (0.09 and 0.14), and for MS the mean allelic error of 0.065 (see text) applied to totals only.

<table>
<thead>
<tr>
<th>Year</th>
<th>TF n</th>
<th>RDF n</th>
<th>LDF n</th>
<th>MS n</th>
</tr>
</thead>
<tbody>
<tr>
<td>1983</td>
<td>2</td>
<td>1</td>
<td>1</td>
<td>–</td>
</tr>
<tr>
<td>1984</td>
<td>2</td>
<td>–</td>
<td>–</td>
<td>–</td>
</tr>
<tr>
<td>1988</td>
<td>3</td>
<td>–</td>
<td>–</td>
<td>–</td>
</tr>
<tr>
<td>1989</td>
<td>1</td>
<td>3</td>
<td>–</td>
<td>–</td>
</tr>
<tr>
<td>1990</td>
<td>3</td>
<td>–</td>
<td>–</td>
<td>–</td>
</tr>
<tr>
<td>1992</td>
<td>2 (1)</td>
<td>2 (2)</td>
<td>1</td>
<td>–</td>
</tr>
<tr>
<td>1993</td>
<td>7</td>
<td>9</td>
<td>9</td>
<td>3</td>
</tr>
<tr>
<td>1997</td>
<td>1</td>
<td>2</td>
<td>3</td>
<td>–</td>
</tr>
<tr>
<td>1999</td>
<td>3</td>
<td>4 (1)</td>
<td>6 (1)</td>
<td>2</td>
</tr>
<tr>
<td>2000</td>
<td>2</td>
<td>5 (2)</td>
<td>7 (2)</td>
<td>5 (2)</td>
</tr>
<tr>
<td>2001</td>
<td>30 (3)</td>
<td>59 (3)</td>
<td>66 (5)</td>
<td>39 (3)</td>
</tr>
<tr>
<td>2002</td>
<td>33 (6)</td>
<td>61 (9)</td>
<td>51 (7)</td>
<td>38 (7)</td>
</tr>
<tr>
<td>2003</td>
<td>24 (7)</td>
<td>38 (8)</td>
<td>35 (10)</td>
<td>29 (7)</td>
</tr>
<tr>
<td>2004</td>
<td>20 (5)</td>
<td>26 (5)</td>
<td>25 (1)</td>
<td>27 (7)</td>
</tr>
<tr>
<td>2005</td>
<td>14 (0)</td>
<td>32 (4)</td>
<td>24 (2)</td>
<td>27 (5)</td>
</tr>
<tr>
<td>2006</td>
<td>24 (4)</td>
<td>31 (4)</td>
<td>32 (4)</td>
<td>22 (6)</td>
</tr>
<tr>
<td>2007</td>
<td>11 (1)</td>
<td>–</td>
<td>–</td>
<td>–</td>
</tr>
<tr>
<td>2008</td>
<td>1 (1)</td>
<td>3 (1)</td>
<td>3 (1)</td>
<td>1</td>
</tr>
<tr>
<td>Totals and (correction)</td>
<td>(none)</td>
<td>154</td>
<td>237</td>
<td>230</td>
</tr>
<tr>
<td>Survival</td>
<td>(survival)</td>
<td>121</td>
<td>189</td>
<td>183</td>
</tr>
<tr>
<td>Survival and Error</td>
<td>(survival and error)</td>
<td>n/a</td>
<td>172</td>
<td>156</td>
</tr>
</tbody>
</table>

TF = tail fluke; RDF = right dorsal fin; LDF = left dorsal fin; MS = microsatellite

### Table 5: Abundance estimates ($N^*$) for humpback whales at Saldanha Bay, WSA, by the Chapman’s modified Petersen method using separate identification features for one pair of capture periods ($j_1 = September 2001–February 2002; j_2 = September 2002–February 2003$). Photographs with quality and orientation ratings of ‘poor’ and lower were excluded from the analysis. Correction factor for dorsal fins refers to calculated false negative rates (RDF = 0.091 and LDF = 0.138, see Table 7), and for microsatellite the mean allelic error rate of 0.065, applied using the method of Stevick et al. (2001) (see text). Percentage bias calculated relative to the uncorrected estimator. (Notation used: n/a = not applicable; $n_i$ = no. of individuals identified during $j_i$ and $n_j$ during $j_j$; $m_j$ = no. of individuals seen in $j_j$ and resighted in $j_j$; SE = standard error; CV = coefficient of variation; LCI = lower 95% confidence intervals; UCI = upper 95% confidence intervals)

<table>
<thead>
<tr>
<th>Feature</th>
<th>Treatment</th>
<th>$n_1$</th>
<th>$n_2$</th>
<th>$m_j$</th>
<th>$N^* \pm SE$</th>
<th>CV ($N^*$)</th>
<th>LCI</th>
<th>UCI</th>
<th>% bias</th>
</tr>
</thead>
<tbody>
<tr>
<td>Tail flukes</td>
<td>Uncorrected</td>
<td>15</td>
<td>16</td>
<td>3</td>
<td>67 ± 23</td>
<td>0.34</td>
<td>35</td>
<td>129</td>
<td>n/a</td>
</tr>
<tr>
<td>Right dorsal fins</td>
<td>Uncorrected</td>
<td>39</td>
<td>58</td>
<td>7</td>
<td>294 ± 82</td>
<td>0.28</td>
<td>172</td>
<td>502</td>
<td>0</td>
</tr>
<tr>
<td></td>
<td>Corrected</td>
<td>38.70</td>
<td>57.30</td>
<td>7.70</td>
<td>265 ± 70</td>
<td>0.26</td>
<td>160</td>
<td>440</td>
<td>10</td>
</tr>
<tr>
<td>Left dorsal fins</td>
<td>Uncorrected</td>
<td>39</td>
<td>49</td>
<td>8</td>
<td>221 ± 56</td>
<td>0.25</td>
<td>136</td>
<td>361</td>
<td>18</td>
</tr>
<tr>
<td></td>
<td>Corrected</td>
<td>38.20</td>
<td>47.08</td>
<td>9.28</td>
<td>182 ± 42</td>
<td>0.23</td>
<td>117</td>
<td>283</td>
<td>18</td>
</tr>
<tr>
<td>Microsatellite</td>
<td>Uncorrected</td>
<td>34</td>
<td>41</td>
<td>9</td>
<td>146 ± 33</td>
<td>0.22</td>
<td>95</td>
<td>225</td>
<td>18</td>
</tr>
<tr>
<td></td>
<td>Corrected</td>
<td>30.58</td>
<td>40.30</td>
<td>9.63</td>
<td>122 ± 25</td>
<td>0.21</td>
<td>81</td>
<td>182</td>
<td>17</td>
</tr>
</tbody>
</table>
lower estimate for N. Model 3 was also fairly well supported for MS data, although model 2 performed better (Table 6); estimates based on this feature showed the least variation between model variants, and the tightest confidence intervals. All other identification features showed considerable variation and very wide CIs, especially those derived from dorsal fins. For all identification features, model 3 — the variant with fewest parameters — yielded the lowest CVs, although it was not well supported (zero likelihood) for the dorsal fin data (Table 6). The estimates for TF (with the exception of model 2) were lower than for other features. The estimates derived from dorsal fins were about double the highest TF or MS estimates, but with high variance and CIs (Table 6). Goodness-of-fit tests available in MARK did not yield results because of inadequate data availability. Given the sparse data and low number of recaptures, no attempt was made to model more complex configurations.

**Potential biases in abundance estimates for different photographic identification features**

**False negatives**

Assuming that the microsatellite identifications were as an identification feature resulted in 13.8% and 9.1% missed matches respectively, whereas no missed matches were detected for tail flukes (Table 7). No false positives were detected for dorsal flunks. To test for misidentifications using microsatellites, individuals resighted by tail flukes on different days using pictures of quality and/or orientation better than poor were used as a control (11 individuals, intercepted 24 times), and were compared to matches obtained by microsatellite (where biopsies were taken). No false negatives were detected in seven matching opportunities. The values for N* for the LDF and RDF recapture data and corrected for by the respective error rates (0.14 and 0.09) were 18% and 10% lower than the respective uncorrected values (Table 5). Although no microsatellite mismatches were detected, an abundance estimate corrected for the mean allelic error rate (0.065) is included for comparison: it was 17% lower than the uncorrected estimate (Table 5).

**Individual variation in fluke exposure relative to other features**

For 21.67% of the whales resighted on different days (n = 60), no pictures of TF were collected, for 20% no biopsies, 3.33% no RDF and 1.67% no LDF photographs. In the

---

**Table 6: Selected parameter estimates and model selection criteria for three model variants in the POPAN version of Jolly-Seber open-population model in MARK 5.1, for different identification features.**

<table>
<thead>
<tr>
<th>Model</th>
<th>N ± SE</th>
<th>CV (N)</th>
<th>LCI</th>
<th>UCI</th>
<th>QAICc</th>
<th>ΔQAICc</th>
<th>Mod. likel.</th>
<th>NP</th>
</tr>
</thead>
<tbody>
<tr>
<td>Tail flukes</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>2 - (\phi_0 \psi_1 \beta_1 \gamma_1 )</td>
<td>531 ± 347</td>
<td>0.65</td>
<td>192</td>
<td>1 771</td>
<td>106.399</td>
<td>0</td>
<td>1</td>
<td>8</td>
</tr>
<tr>
<td>3 - (\phi_0 \psi_2 \beta_2 \gamma_2 )</td>
<td>301 ± 100</td>
<td>0.33</td>
<td>172</td>
<td>588</td>
<td>106.587</td>
<td>0.1880</td>
<td>0.9103</td>
<td>4</td>
</tr>
<tr>
<td>1 - (\phi_0 \psi_3 \beta_3 \gamma_3 )</td>
<td>233 ± 113</td>
<td>0.48</td>
<td>116</td>
<td>620</td>
<td>112.112</td>
<td>5.7127</td>
<td>0.0575</td>
<td>11</td>
</tr>
<tr>
<td>Microsatellites</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>2 - (\phi_0 \psi_4 \beta_4 \gamma_4 )</td>
<td>528 ± 143</td>
<td>0.27</td>
<td>332</td>
<td>922</td>
<td>230.686</td>
<td>0</td>
<td>1</td>
<td>9</td>
</tr>
<tr>
<td>3 - (\phi_0 \psi_5 \beta_5 \gamma_5 )</td>
<td>400 ± 65</td>
<td>0.16</td>
<td>301</td>
<td>561</td>
<td>234.518</td>
<td>3.8323</td>
<td>0.1472</td>
<td>2</td>
</tr>
<tr>
<td>1 - (\phi_0 \psi_6 \beta_6 \gamma_6 )</td>
<td>496 ± 145</td>
<td>0.29</td>
<td>305</td>
<td>907</td>
<td>235.006</td>
<td>4.3198</td>
<td>0.1153</td>
<td>11</td>
</tr>
<tr>
<td>Right dorsal fins</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>2 - (\phi_0 \psi_7 \beta_7 \gamma_7 )</td>
<td>1 035 ± 375</td>
<td>0.36</td>
<td>552</td>
<td>2 117</td>
<td>198.517</td>
<td>0</td>
<td>1</td>
<td>7</td>
</tr>
<tr>
<td>1 - (\phi_0 \psi_8 \beta_8 \gamma_8 )</td>
<td>955 ± 495</td>
<td>0.52</td>
<td>420</td>
<td>2 604</td>
<td>206.221</td>
<td>7.7041</td>
<td>0.0212</td>
<td>11</td>
</tr>
<tr>
<td>3 - (\phi_0 \psi_9 \beta_9 \gamma_9 )</td>
<td>681 ± 139</td>
<td>0.20</td>
<td>472</td>
<td>1 032</td>
<td>231.399</td>
<td>32.8820</td>
<td>0</td>
<td>3</td>
</tr>
<tr>
<td>Left dorsal fins</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>1 - (\phi_0 \psi_10 \beta_{10} \gamma_{10} )</td>
<td>1 232 ± 774</td>
<td>0.63</td>
<td>455</td>
<td>3 951</td>
<td>141.953</td>
<td>0</td>
<td>1</td>
<td>11</td>
</tr>
<tr>
<td>2 - (\phi_0 \psi_11 \beta_{11} \gamma_{11} )</td>
<td>1 013 ± 498</td>
<td>0.49</td>
<td>449</td>
<td>2 612</td>
<td>146.357</td>
<td>4.4036</td>
<td>0.1106</td>
<td>7</td>
</tr>
<tr>
<td>3 - (\phi_0 \psi_12 \beta_{12} \gamma_{12} )</td>
<td>760 ± 194</td>
<td>0.26</td>
<td>481</td>
<td>1 270</td>
<td>178.347</td>
<td>36.3940</td>
<td>0</td>
<td>2</td>
</tr>
</tbody>
</table>

---

**Table 7: False-negative rates (number of missed matches as % of total number of matching opportunities) detected for humpback whale photographic identification features, west South Africa, using microsatellite matches as a control.**

<table>
<thead>
<tr>
<th>ID feature</th>
<th>Sample occasions</th>
<th>Matching opportunities</th>
<th>Confirmed matches</th>
<th>Missed matches</th>
<th>False-negative rate (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>MS (control)</td>
<td>88</td>
<td>32</td>
<td>–</td>
<td>–</td>
<td>–</td>
</tr>
<tr>
<td>LDF</td>
<td>58</td>
<td>29</td>
<td>25</td>
<td>4</td>
<td>13.8</td>
</tr>
<tr>
<td>RDF</td>
<td>49</td>
<td>22</td>
<td>20</td>
<td>2</td>
<td>9.09</td>
</tr>
<tr>
<td>TF</td>
<td>30</td>
<td>13</td>
<td>13</td>
<td>0</td>
<td>0</td>
</tr>
</tbody>
</table>
majority of cases, TF photographs (for the 47 whales) were obtained during the first intercept/encounter (65.96%), 27.66% during the second and 6.38% during the third and fourth. Furthermore, during all intercepts involving these resighted whales \((n = 183\); some whales were in the same groups), TF pictures were collected during only 57.4% of intercepts, compared to 92.9% for dorsal fins. There was no significant difference between the mean duration of intercepts where TF were photographed \((73.84 \text{ min, SE } 3.88, n = 146)\) and where they were not \((83 \text{ min, SE } 11.14, n = 31)\) \((t = -0.93, df = 175, p = 0.35)\). The probability of recording a dorsal fin image (right or left) of an individual whale every time it was encountered was high (Figure 6). This was not the case for TF, where for individual whales the probability of recording this feature during all, half or none of encounters was very similar (28%, 25% and 23% respectively) (Figure 6).

**Double-mark models**

For the selected capture periods used for CMP estimates, the models where TF were used for recapture \((m_3)\) and the second sample \((n_2)\) invariably resulted in lower abundance estimates than when the alternative features were used (Table 8). The highest estimates were calculated with RDF as the alternative mark, while LDF and MS yielded very similar estimates. Those in which TF were used for \(n_2\) were 0.31, 0.39 and 0.43 of those using RDF, LDF and MS respectively for \(n_2\).

**Discussion**

**Sighting database, resightings and migratory links**

The distribution over space and time of contributions to the sighting database, and resulting resighting rates, reflected the generally low and often inconsistent collection effort. This makes it difficult to interpret the observed resighting rates relative to other capture–recapture studies with greater geographic coverage and higher sampling effort, such as is obtainable during dedicated survey cruises (e.g. Larsen and Hammond 2004, Wedekin et al. 2010) or simultaneous surveys from multiple platforms or sites (e.g. Smith et al. 1999, Calambokidis and Barlow 2004). Resighting rates are known to be much higher at feeding sites (some >50%) compared to breeding grounds (10% or less) (Herman et al. 2011). Our between-year resighting rate of 15.65% seems relatively high given the low effort, limited extent of sampling, and that it is neither a breeding nor typical feeding area. This could thus be indicative of strong site fidelity or a small ‘population’, or both. There is strong historic evidence for severe depletion of this assemblage of humpback whales: during the five years immediately prior to protection of the species in 1963, annual catches from the Donkergat whaling station at Saldanha Bay averaged only five whales taken during a six-month season (BIWS 1964), compared to catches of 208–244 humpback whales a season over the first three years of whaling from Donkergat (Best 1994).

Long-term site fidelity is supported by the majority of individual resightings occurring at annual or biennial intervals, and on six occasions up to a decade apart. While most of these whales were seen only twice, others were seen in three or more different calendar years up to a maximum of six different years; again, the ad hoc collection effort probably confounded the ability to detect more returns of known animals. Returns over such time-scales may confirm fidelity to the area but do not necessarily identify it as anything other than a migratory corridor. However, this is challenged by sequential resightings of the same individuals on the same day, or within a week of the first sighting, suggesting that they were not merely moving

![Figure 6: Probability of recording a photographic identification feature for 60 individual (resighted) humpback whales off west South Africa, calculated as the number of times a feature (tail fluke or dorsal fin) was recorded as the proportion of the total number of times that the whale was intercepted](image)

Table 8: Abundance estimates \((N^*)\) for west South African humpback whales from the Chapman’s modified Petersen estimator for various model configurations using double marked (TF plus alternative mark) humpback whales identified during the first sampling period, and recaptures based on TF or alternative mark during the second sampling period. Notation used: SE = estimated standard error, CV = estimated coefficient of variation, LCI and UCI = lower and upper 95% confidence intervals

<table>
<thead>
<tr>
<th>Model</th>
<th>(n_1)</th>
<th>(n_2^*)</th>
<th>(m_2^*)</th>
<th>(N^*)</th>
<th>SE((N^*))</th>
<th>CV((N^*))</th>
<th>LCI</th>
<th>UCI</th>
</tr>
</thead>
<tbody>
<tr>
<td>(n_1 = \text{TF&amp;RDF}, n_2 = \text{RDF, } m_2 = \text{RDF})</td>
<td>10</td>
<td>57.30</td>
<td>3.30</td>
<td>148</td>
<td>48.65</td>
<td>0.33</td>
<td>79</td>
<td>277</td>
</tr>
<tr>
<td>(n_1 = \text{TF&amp;RDF, } n_2 = \text{TF, } m_2 = \text{TF})</td>
<td>10</td>
<td>7.26</td>
<td>3</td>
<td>46</td>
<td>14.58</td>
<td>0.32</td>
<td>25</td>
<td>84</td>
</tr>
<tr>
<td>(n_1 = \text{TF&amp;LDF, } n_2 = \text{LDF, } m_2 = \text{LDF})</td>
<td>11</td>
<td>47.08</td>
<td>3.48</td>
<td>128</td>
<td>41.47</td>
<td>0.32</td>
<td>69</td>
<td>238</td>
</tr>
<tr>
<td>(n_1 = \text{TF&amp;RDF, } n_2 = \text{TF, } m_2 = \text{TF})</td>
<td>11</td>
<td>16</td>
<td>3</td>
<td>50</td>
<td>16.28</td>
<td>0.33</td>
<td>27</td>
<td>93</td>
</tr>
<tr>
<td>(n_1 = \text{TF&amp;MS, } n_2 = \text{MS, } m_2 = \text{MS})</td>
<td>9</td>
<td>40.30</td>
<td>2.14</td>
<td>131</td>
<td>51.50</td>
<td>0.39</td>
<td>62</td>
<td>275</td>
</tr>
<tr>
<td>(n_1 = \text{TF&amp;MS, } n_2 = \text{TF, } m_2 = \text{TF})</td>
<td>9</td>
<td>16</td>
<td>2</td>
<td>56</td>
<td>21.51</td>
<td>0.39</td>
<td>27</td>
<td>116</td>
</tr>
</tbody>
</table>

* An error correction of 0.065 for MS, 0.09 for RDF, and 0.14 for LDF was applied for \(n_2\) and \(m_2\)
through the area as expected during a typical migration. This confirms the phenomenon of temporary residency first observed during the 1993 study at Cape Columbine (Best et al. 1995), when the same 10 humpback whales were seen on average 2.4 times during a period of 1.5 months. It is thus not inconceivable that whales resighted in the same year or breeding season at periods of 1–6 months apart could be moving around locally and remain in the general area for days, weeks or even months. Continuous occupancy cannot be proven, however, and it should be equally feasible for animals to depart from and return to the area in such time-spans. Here, the matches with the two other regional catalogues shed more light on the possible nature of such movements. The resightings off Cape Town detected through the AHWCM matches confirm the occurrence of local movements beyond the core study area, and a net southward movement during summer. The three matches between Gabon and WSA independently confirm evidence from genotypic matches (Carvalho et al. 2010) that showed exchange of individual whales between these two proposed ‘substocks’. All three records indicate movement between winter (August, September) in Gabon and late spring/summer (November–February) in WSA, with the movement occurring in both directions, but with the fastest transits from north to south. The monthly distribution of animals that were seen on more than one occasion (both within- and between-region resightings) suggests that humpback whales that engage in feeding during late spring, and in particular the summer months off the west coast of South Africa (Barendse et al. 2010), are also likely to be encountered repeatedly during these months in other years. Furthermore, some of these whales were present off Gabon during August/September, so presumably overwintered there. On the other hand, the paucity of resightings of any of the animals identified off WSA during June–August (mid-winter), during which 11.38% of all whales identified were seen (based on combined features), suggests that whales present in the region at that time might belong to a different component of the population. If, as suggested by other evidence (see Chittleborough 1965, Dawbin 1966), this corresponds to the timing of an expected northern migration, it would appear that animals utilising the study area as a spring/summer feeding ground on their southward migration do not necessarily take the same route when moving north to the breeding grounds.

The lack of any matches with the few available Namibian animals does not preclude the occurrence of a ‘typical’ coastal migration from headland to headland (Chittleborough 1965, Dawbin 1966) from WSA through Namibia to a more northerly destination, and return on the southern migration. However, historical evidence does not support this notion, in that, off Namibia, catches showed a sign of ‘recovery’ during the period 1925–1930 after initial depletion, while off WSA they remained very low (Best and Allison 2010). Possibly the coastal migration stream is cumulative rather than unitary, with northward-moving animals progressively converging on the coast with decreasing latitude, and southward-moving animals leaving the coast with increasing latitude. At Walvis Bay there were some recent sightings during late summer, but most humpback whale sightings were in winter (June, July, August) (SHE unpublished data). However, there was no research effort after these months to allow detection of a later peak, as observed off WSA, although whale- and dolphin-watching operators did encounter some humpback whales during September and October. Furthermore, historical catches (Best and Allison 2010) showed strong bimodality at Walvis Bay, with a peak in June/July and another in October/November.

**Abundance estimates**

The available capture–recapture data were very limited in terms of sample sizes and the number of recaptures detected between sampling periods. This was especially so for TF which were under-represented in the database with only 121 individuals (after correction for survival), while the similarly low number of whales identified by MS could be partly attributed to its implementation (as a means of identification) a decade later than photo-identification. The higher numbers of whales identified by dorsal fins may reflect the fact that they are more easily photographed: unlike TF, they are always exposed during surfacings (but see below), whereas biopsy sampling requires the closest approach of all sampling methods and may be difficult to achieve for boat-shy individuals that could still be photographed. The CMP estimates for $N^*$ during the first sampling periods more or less reflect this relative representation of features in the sighting database, i.e. dorsal fins the highest and TF the lowest at an improbable 67 individuals. For the remaining features, the estimates for $N^*$ ranged between 122 (MS) and 265 (RDF) after error correction. That the TF estimate is an order of magnitude lower than for any other feature and is comparable to the adjusted catalogue size for TF at this time (69), suggests that the ability to capture a whale using TF may be affected by an additional factor (see below).

The generally poor (or non-) performance of the POPAN open-population models when capture probability ($p$) and probability of entry ($β$) were fixed again suggests considerable variation in these parameters between capture periods. This is perhaps unsurprising, given the low effort, the differences in sampling strategy (with or without a land watch) and location of sampled areas (Saldanha vs St Helena Bay) between $j_1–j_2$ and $j_3–j_6$. For the most successful model variants, apparent survival ($ϕ$) was fixed at 0.96 and both $β$ and $p$ were fully time-dependent (model 2), or $β$ time-dependent and $p$ fixed (model 3). Before looking at the magnitude of the estimated values for $N$ (the super-population) by the POPAN models, it is worth considering it in the context of what was sampled. This parameter provides an estimate for the total number of animals, both captured or not, available in a (hypothetical?) super-population (GC White, Program MARK Help files). For example, in a study of bottlenose dolphins, Reisinger and Karczmarski (2010) using POPAN concluded that the $N$ potentially represented the dolphin population along a considerable segment of (or even the entire) South African coastline, whereas in other studies it has been taken to represent a full population of right whales (Wade et al. 2011), a subpopulation of whale sharks (Meekan et al. 2006) or a feeding assemblage of humpback whales (Larsen and Hammond 2004). Thus, it is apparent that $N$, as estimated by POPAN, is likely to represent more than simply the size of the feeding assemblage at Saldanha and St Helena Bays during any given season, but probably
also includes whales that migrated through the area, or whose use of the area varied between years, and should probably be treated as an upper subpopulation limit.

The abundances obtained from the open-population models were larger than those using other methods. Similar to the other assessment methods, the dorsal fin estimates were twice as high as those for other features, most likely as a result of the occurrence of false-negative errors (see below). Although a correction parameter for photographic and genotypic mismatches is not available in the POPAN model, estimates that include such errors would most likely still fall within the wide confidence intervals obtained, especially for dorsal fins. Tail fluke estimates were lower than for other features (but not as markedly as with the CMP model), with the exception of model 2 in which the estimate was very similar to that for MS, albeit with a much higher CV. The MS capture–recapture data yielded the most consistent estimates with the lowest variation between models, and tightest confidence intervals for individual variants. The model-averaged estimate of N for this feature (calculated from all three variants in MARK) was 510 (SE 143; 95% CIs = 230–790).

**Biases as a result of different identification features**

There are numerous known issues associated with the use of natural markings for abundance estimates (see Hammond 1986 for review), but (physical) loss of marks is not considered to be a major problem with humpback whales. Dorsal fins are commonly used by researchers in the field to distinguish between different whales in a group while collecting data during boat encounters, and they have been proposed as a potentially even more stable identification feature than ventral tail fluke pigmentation (Blackmer et al. 2000). To our knowledge, however, our study is the first in which dorsal fin markings have been used to provide an abundance estimate for humpback whales. While false-positives are probably rare in photo-identification studies (we detected none), false-negatives are thought to be more common; the poorer the quality of pictures, the higher the error rate (Stevick et al. 2001). This is more likely to apply to dorsal fins as they are smaller and have fewer distinguishing features than tail flakes. Although dorsal fin photographs (of sufficient quality) were not collected during all intercepts, there were no resighted individuals for which dorsal fin pictures were unavailable. There may be differences in the ability of researchers to obtain good quality images of these different identification features: during a typical approach from the rear, chances are good of obtaining a TF picture (provided that they are adequately exposed, see below). For dorsal fins, a considerable amount of manoeuvring of the boat is required to position the photographer at a right angle to a surfacing whale. The angle between the camera and the whale affects the quality of dorsal fin pictures to a greater extent than for TF (JB pers. obs.). Poor image quality can be the source of substantial heterogeneity in capture probability when using dorsal fins as identification features in other species (e.g. northern bottlenose whales *Hyperoodon ampullatus*; Gowans and Whitehead 2001). The application of some quality control (e.g. removal of photos with incomplete subjects and those of low quality) has been shown to adequately reduce error rates when using TF (Friday et al. 2008), but it is unknown to what degree this is applicable to dorsal fins, and we were unable to test this with such small sample sizes.

Assuming the microsatellite identifications were correct, dorsal fin photographs used alone as an identification feature resulted in 9–14% missed matches, whereas this does not appear to apply to the same extent for TF. If left uncorrected, this may result in a substantial overestimation (up to 30%) of abundance when using closed-population models, and produce high estimates of low confidence in open-population models. This conclusion, however, may be case-specific, depending to a large extent on data collection protocol, photographic quality, laboratory procedures, and the size of the catalogue. The differences between abundance estimates for RDF and LDF (although less pronounced compared to TF) suggests that there may have also been a difference in the ability of photographers to obtain usable images from both sides of an individual. The reason for this is not immediately apparent, although individual whale behaviour could contribute to such a bias. Clapham et al. (1995) reported strongly lateralised behaviour by humpback whales that apparently favoured their right side during feeding and flippering behaviour; it is possible that whales could preferentially present their right side to the boat. However, we are unable to test this with the available data.

Relative to other identification features (even after they had been corrected for missed matches), TF yielded the overall lowest abundance estimates. This suggests that fluking as an individual behavioural trait could affect the probability of an individual being sampled, and was supported by the finding that for resighted whales, the probability of collecting TF pictures during all, half or none of the intercepts was nearly equal. The fact that for all resighted whales, over 20% had no TF image collected at all during intercepts of similar mean duration, and that in the majority of cases (65%) flukes were photographed during the first intercept, suggests that fluking is an idiosyncratic feature for humpback whales in this area. The frequency of exposure of the ventral surfaces of the flukes is a behaviour known to vary with sex (Rice et al. 1987), age, reproductive and behavioural class, and group size, with fluking rates ranging from <10 per 100 dives for cows, calves and principal escorts, to 46.5 per 100 dives for single animals (Smith et al. 1999). If some whales consistently fluke less often than others, or do not fluke at all, the resulting heterogeneity of capture probabilities will lead to underestimation of population size (Barlow et al. 2011), such as is strongly suggested by these data. In West Greenland, such (downward) bias was estimated at 10–20% of the population size (Perkins et al. 1984), but presumably can vary with area, season or photographic protocol. Based on the CMP abundance estimates for the double-marked whales, those using TF recaptures and identifications during the second sampling period were 57–69% lower than those using an alternative feature. While this conclusion about the effects of individual fluking behaviour on population estimation may only be valid for the whales observed in some areas (such as WSA), as humpback whale behaviour may differ (and sampling protocol vary) in different parts of their range, the potential effects shown here are certainly large enough to
warrant similar investigations in other areas. It is, however, more difficult to quantify (and thus correct for) heterogeneity attributable to individual behaviour compared to other sources of error (e.g. photographic quality) (Barlow et al. 2011).

Genotypic abundance estimates may be considered independent from those obtained from photo-identification (as suggested by Gubili et al. 2009) with a lesser degree of bias from sampling heterogeneity applicable to photographs of natural marks (i.e. image quality and fluking behaviour). On a broader scale, because genetic and photographic sampling took place simultaneously and from the same platform, both could be considered subject to the same potential biases caused by non-representative sampling effort. The abundance estimates derived from MS recaptures were lower than those from dorsal fins, but similar (although higher) to those obtained for TF, though more recaptures were made with MS. Heterogeneity of obtaining a biopsy (i.e. capture probability) can therefore not be excluded, given that it requires a closer approach than obtaining a TF image, and recognising that for 20% of resighted whales no biopsy was collected. This could be as a result of a different individual behavioural response to boat approaches, as has been tested for whale-watching boats (e.g. Scheidat et al. 2004, Stamation et al. 2010): the whales could exhibit boat avoidance (i.e. ‘trap-shy’ animals) or be boat-friendly (‘trap-happy’), both of which will cause capture heterogeneity and bias in abundance estimates. Moreover, there are certain issues relating to methodology and laboratory procedures that are specific to the use of molecular tags and may bias abundance estimates downward (Mills et al. 2000, Waits and Leberg 2000) or upward (Lukacs and Burnham 2005, Wright et al. 2009).

Conclusion

The consolidated photographic and genotypic sighting database for humpback whales from WSA contributes to a better understanding of residency rates and long-term attendance patterns in the region. It reveals that some whales routinely visit the coastal waters of the St Helena/ Saldanha Bay region, showing high fidelity to a geographically limited area, to participate in feeding during spring and summer. While acknowledging the effects of the sampling approach, low effort and small sample sizes on capture probability, possible structuring of the population would further contribute heterogeneity to individual capture, and the notion that there may be a strictly migratory (or ‘transient’) component is supported by the high number of once-off sightings, especially during winter. The situation is possibly fairly complex, but similar to that observed offshore California where whales that share a wintering region show strong fidelity to specific feeding areas, with limited exchange between these, although whales from different subareas may make use of the same migratory corridors (Calambokidis et al. 2001). Given the limitations of the data, and violation of closure, the estimates from the closed-population model are likely to be underestimates of the total number of humpback whales found here on a seasonal basis, although their use did allow us to explore issues related to the use of different identification features. The findings that using dorsal fins and TF can cause substantial over- and underestimation respectively are important, especially when dealing with such a small dataset, and given that the ventral TF pattern is the standard identification feature used for humpback whales. Whether humpback whale dorsal fins are distinctive enough features to use in large catalogues (or as an alternative identification feature) is debatable. It may be more appropriate to only include animals that are considered adequately ‘marked’, similar to the practise in dolphin studies (e.g. Elwen et al. 2009), although this approach may violate assumptions of equal capture probability, depending on the model used (see Reisinger et al. 2011). The open-population models with few restrictions and using multiple-capture periods fitted the capture–recapture data better, although there was considerable variation between the different identification features and model variants used. The most consistent estimates (and those with the tightest confidence intervals) were obtained from the microsatellite data, which put the number of humpback whales that visited the area during the study period at about 500 animals, a value that falls within the confidence intervals for dorsal fins (lower range) and tail flukes (mid–upper range). The geographic extent of this estimate is not clear, nor whether all of these animals feed in the region of Saldanha every season, or how they relate to whales along the rest of the West African coast. Although this assemblage does not exist in isolation from the greater Breeding Stock B (given the photographic matches), it does seem to represent a previously undescribed situation in the Southern Hemisphere. However, the possibility cannot be excluded that such behaviour or similar assemblages may occur at other places where comparable oceanographic conditions (i.e. wind-driven upwelling) are present. For example, some Namibian localities such as Lüderitz (Hutchings et al. 2009) may have similar feeding opportunities to those off WSA. For a better understanding of the population structure in the region, research effort and photographic and genetic data collection need to cover more seasons, and include more sites within WSA and the rest of West Africa — including farther offshore — similar to some of the long-term, ocean-basin wide identification studies such as the SPLASH (Structure of Populations, Levels of Abundance, and Status of Humpbacks) programme in the North Pacific (Calambokidis et al. 2008), or YoNAH (Year of the North Atlantic Humpback; Smith et al. 1999). The strategic deployment of satellite tags off WSA may offer a more short-term solution to elucidate potential migratory routes and locations of unknown breeding or feeding areas (e.g. Zerbini et al. 2006, Hauser et al. 2010), which would help inform whether the current IWC management units are relevant to humpback whales in this region.

Acknowledgements — Research work during the main research project off WSA was supported by the National Research Foundation (NRF), South Africa, under Grant Number 2047517. The following other organisations are thanked: Earthwatch Institute (funding); the Mazda Wildlife Fund (through the provision of a field vehicle); SASOL (through the donation of two four-stroke engines); PADI Project AWARE (UK) (funding); the South African Navy (access to the shore-based look-out); the Military Academy, University of Stellenbosch (accommodation); and Iziko South African Museum (office space and support). JB gratefully received
financial support in the form of bursaries from the NRF, the Society for Marine Mammalogy, University of Pretoria, and the Wildlife Society of South Africa (Charles Astley Maberley Memorial bursary). The Namibian Dolphin Project is supported by the Namibian Coastal Conservation and Management Project, the Nedbank Go Green Fund, Mohammed bin Zayed Species Conservation Fund, the British Ecological Society, the Rufford Small Grants Foundation, and the Namibia Nature Foundation. JB and TJQC received funding from the International Whaling Commission to conduct between-region matching. CP, IC and HCR are grateful to the Sakeller Institute for Comparative Genomics, American Museum of Natural History, in particular Dr George Amato, Dr Rob DeSalle, Matt Leslie and Jacqueline Ay-Ling Loo. The fieldwork would have been impossible without the enthusiastic assistance of numerous Earthwatch and other volunteers, to all of whom we owe a big debt of gratitude. The following photographers (from various affiliations) took or contributed images in RSA: Blake Abernethy, Meredith Thornton, Ingrid Peters, Desray Reeb, Shaun Dillon, Lisa Mansfield, Tilen Genov, Stephanie Plön, André du Randt, Mike Meijer, Darelle Anders, Sharon du Plessis, and Rob Tarr; and in Namibia: Mike Lloyd, Ute von Ludwiger, Orlanda Sardinha and Francois Visser (Levo Tours). The scanning protocol and photographic database structure was based on one developed by PJ Ersts (later modified by S Cerchio, TJQC and WCS) as part of a regional Atlantic/Indian Ocean humpback whale research collaboration. South African digital coastline data were obtained from the South African National Biodiversity Institute as used for the National Spatial Biodiversity Assessment 2004. Ocean-wide bathymetry shape files were downloaded from Natural Earth (http://www.naturalearthdata.com/). Steve Kirkman, Ingrid Peters and Salvatore Cerchio are thanked for discussions on mark-recapture analyses. An earlier version of the manuscript benefitted greatly from the comments of two anonymous referees. Research in South Africa was authorised by successive annual permits issued to PBB by the Minister for Environmental Affairs, in terms of Regulation 58 of the Marine Living Resources Act, 1998 (Act No. 18 of 1998). Research in Namibia was completed under a permit issued to SE and RL by the Namibian Ministry of Fisheries & Marine Resources. Some of these data and analyses have previously been presented to the IWC Scientific Committee Meeting in Agadir, Morocco, in 2010, and were included in an unpublished report to this meeting; this report can be accessed at http://iwcoffice.org/_documents/sci_com/SC62docs/SC-62-SH2.pdf.

References


Collins TJQ, Cerchio S, Pomilla C, Loo J, Carvalho I, Ngouesso S, Rosenbaum HC. 2008. Revised estimates of abundance for


Appendix: Summary capture–recapture statistics


| $m_j$ | $j_1$ | $j_2$ | $j_3$ | $j_4$ | $j_5$ | $j_6$ | $m_j$ | $j_1$ | $j_2$ | $j_3$ | $j_4$ | $j_5$ | $j_6$
|-------|-------|-------|-------|-------|-------|-------|-------|-------|-------|-------|-------|-------|-------|-------|-------|-------|-------|
| Tail flukes | Right dorsal fins | Microsatellites
<table>
<thead>
<tr>
<th></th>
<th></th>
<th></th>
<th></th>
<th></th>
<th></th>
<th></th>
<th></th>
<th></th>
<th></th>
<th></th>
<th></th>
<th></th>
<th></th>
<th></th>
<th></th>
<th></th>
<th></th>
</tr>
</thead>
<tbody>
<tr>
<td>$j_1$</td>
<td>x</td>
<td>3</td>
<td>1</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>$j_1$</td>
<td>x</td>
<td>7</td>
<td>1</td>
<td>2</td>
<td>1</td>
<td>2</td>
<td>$j_1$</td>
<td>x</td>
<td>9</td>
<td>2</td>
</tr>
<tr>
<td>$j_2$</td>
<td>–</td>
<td>x</td>
<td>0</td>
<td>1</td>
<td>0</td>
<td>1</td>
<td>$j_2$</td>
<td>–</td>
<td>x</td>
<td>0</td>
<td>4</td>
<td>1</td>
<td>1</td>
<td>$j_2$</td>
<td>–</td>
<td>x</td>
<td>0</td>
</tr>
<tr>
<td>$j_3$</td>
<td>–</td>
<td>–</td>
<td>x</td>
<td>1</td>
<td>0</td>
<td>0</td>
<td>$j_3$</td>
<td>–</td>
<td>–</td>
<td>x</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>$j_3$</td>
<td>–</td>
<td>–</td>
<td>x</td>
</tr>
<tr>
<td>$j_4$</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>x</td>
<td>0</td>
<td>0</td>
<td>$j_4$</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>x</td>
<td>1</td>
<td>0</td>
<td>$j_4$</td>
<td>–</td>
<td>–</td>
<td>–</td>
</tr>
<tr>
<td>$j_5$</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>x</td>
<td>1</td>
<td>$j_5$</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>x</td>
<td>0</td>
<td>$j_5$</td>
<td>–</td>
<td>–</td>
<td>–</td>
</tr>
<tr>
<td>$m$</td>
<td>0</td>
<td>3</td>
<td>1</td>
<td>2</td>
<td>0</td>
<td>2</td>
<td>$m$</td>
<td>0</td>
<td>7</td>
<td>1</td>
<td>6</td>
<td>3</td>
<td>3</td>
<td>$m$</td>
<td>0</td>
<td>9</td>
<td>58</td>
</tr>
<tr>
<td>$n$</td>
<td>15</td>
<td>16</td>
<td>10</td>
<td>7</td>
<td>9</td>
<td>16</td>
<td>$n$</td>
<td>39</td>
<td>58</td>
<td>14</td>
<td>20</td>
<td>25</td>
<td>27</td>
<td>$n$</td>
<td>39</td>
<td>51</td>
<td>13</td>
</tr>
<tr>
<td>$u$</td>
<td>15</td>
<td>13</td>
<td>9</td>
<td>5</td>
<td>9</td>
<td>14</td>
<td>$u$</td>
<td>39</td>
<td>51</td>
<td>13</td>
<td>14</td>
<td>22</td>
<td>24</td>
<td>$u$</td>
<td>39</td>
<td>51</td>
<td>13</td>
</tr>
<tr>
<td>$M$</td>
<td>0</td>
<td>15</td>
<td>28</td>
<td>37</td>
<td>42</td>
<td>51</td>
<td>$M$</td>
<td>0</td>
<td>39</td>
<td>90</td>
<td>103</td>
<td>117</td>
<td>139</td>
<td>$M$</td>
<td>0</td>
<td>39</td>
<td>90</td>
</tr>
</tbody>
</table>